



UNIVERSITI PUTRA MALAYSIA

**NITROUS OXIDE EMISSION FROM AN UPLAND CROPPING
SYSTEM IN THE HUMID TROPICS**

MOHAMMAD IBRAHIM KHALIL

FP 2001 28

**NITROUS OXIDE EMISSION FROM AN UPLAND CROPPING SYSTEM
IN THE HUMID TROPICS**

BY

MOHAMMAD IBRAHIM KHALIL

**Thesis Submitted in Fulfilment of the Requirement for the Degree of
Doctor of Philosophy in the Faculty of Agriculture
Universiti Putra Malaysia**

March 2001



To My Respected Parents
Shamina Khatun and Late Munshi Sultan Ahmed
and
Beloved Wife, Lucky
Dearest Daughter, Chaity

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

**NITROUS OXIDE EMISSION FROM AN UPLAND CROPPING SYSTEM
IN THE HUMID TROPICS**

By

MOHAMMAD IBRAHIM KHALIL

March 2001

Chairman: Associate Professor Dr. Rosenani Abu Bakar

Faculty: Agriculture, Universiti Putra Malaysia

Co-Chairman: Professor Dr. Ir. Oswald Van Cleemput

Faculty: Agriculturual & Applied Biological Sciences, Ghent Universi

Nitrous oxide (N_2O) emission to the atmosphere has a great implication on global climate change. Agricultural soils seem to be its major source, though little attention is given to the soils and upland cropping systems of the humid tropics. Thus, laboratory experiments were carried out to study the impact of N sources, moisture regimes and soil types on N_2O production. A field experiment was conducted to measure N_2O emissions from a maize-groundnut cropping system managed with different N sources. The laboratory incubation study using an Ultisol showed a maximum N_2O flux of $2379 \mu\text{g N}_2\text{O-N kg}^{-1} \text{ soil d}^{-1}$ with chicken manure application at 60% water-filled pore space (WFPS). Application of potassium nitrate, groundnut residue and urea resulted in smaller production rates ($615 - 699 \mu\text{g N}_2\text{O-N kg}^{-1} \text{ soil d}^{-1}$). Addition of ammonium sulfate and maize residue produced the lowest rates, 229 and $246 \mu\text{g N}_2\text{O-N kg}^{-1} \text{ soil d}^{-1}$, respectively. In general, the total N_2O production in 25 days increased with decrease in C/N ratio of the organic N sources. The loss of

applied N through N₂O emission was higher from inorganic N (3.5-8.6%) than from organic N sources (1.6-6.7%). It could be because of denitrification during the initial period of incubation with readily available mineral N, compared to slower release from organic N sources. Although smaller N₂O production (26.6-38.7 µg N₂O-N kg⁻¹ soil d⁻¹), the fluxes increased with increase in soil moisture content. The relatively drier soil (20% WFPS) acted as a sink. The total N₂O production in the soil with 40, 60 and 80% WFPS increased by 46, 58 and 72%, respectively over the soil with 20% WFPS. Liming the acid soils, similar to the addition of urea and chicken manure, increased the soil pH to around 5.5, stimulating nitrate accumulation after a lag period and N₂O production concurrently. The N₂O productions were not affected by the soil acidity; the total production correlated positively with pH, CEC, organic C and N content of the soils and negatively with water-soluble organic carbon (WSOC). Under the maize-groundnut crop rotation, addition of chicken manure resulted in a maximum N₂O flux of 9889 µg N₂O-N m⁻² d⁻¹ within the first one week after application during the fallow period i.e. after the groundnut crop cycle. The residual effect is also exhibited during the maize cultivation, showing a higher N₂O flux (4053 µg N₂O-N m⁻² d⁻¹) than the plots treated with only inorganic N fertilizer. A lower N₂O flux or negative flux during fallow periods occurred probably due to small availability of substrates and/or low WFPS (<40%). The added N sources retained in the soil for 2 to 3 weeks, matching with the N₂O emission. The high coefficients of variation of N₂O emission under both crop covers showed no clear diurnal variations of N₂O flux. The temporal variability was large, showing a higher emission during the fallow period after addition of chicken manure as well as during maize cultivation after application of N fertilizer. The highest total emission (1.82 kg N₂O-N ha⁻¹) during maize period was in the plots with chicken manure and addition of half the

amount of recommended N fertilizer. This depicted an influence of chicken manure, which was applied before cultivation of the maize crop. The estimated annual emission was 3.94, 1.90 and 1.41 kg N₂O-N ha⁻¹ from the plots treated with chicken manure plus crop residues and N fertilizer, crop residues and N fertilizer, and N fertilizer only, respectively. The estimated fertilizer-induced N₂O emission factor (1.06%) was lower than the generally accepted standard value (1.25%) currently being used by the Intergovernmental Panel on Climate Change. This study suggests that supply of chicken manure to crop fields could be an important potential source of N₂O.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PEMERUWAPAN NITRUS OKSIDA DARIPADA SISTEM PENANAMAN
TANAH TINGGI DI KAWASAN LEMBAB TROPIKA**

Oleh

MOHAMMAD IBRAHIM KHALIL

Mac 2001

Pengerusi: Profesor Madya Dr. Rosenani Abu Bakar

Fakulti: Pertanian, Universiti Putra Malaysia

Pengerusi-bersama: Profesor Dr. Ir. Oswald Van Cleemput

Fakulti: Pertanian dan Biologi Sains Gunaan, Ghent Universiti

Nitrus oksida (N_2O) mempunyai implikasi yang besar terhadap perubahan cuaca global. Tanah pertanian merupakan punca utama pengeluaran N_2O walaupun masih kurang perhatian diberikan terhadap tanah dan sistem pananaman di kawasan tropika. Berdasarkan permasalahan tersebut kajian di ladang dan makmal telah dijalankan untuk mengkaji kesan sumber N, kelembapan tanah dan jenis-jenis tanah terhadap penghasilan N_2O dan pemeruwapannya daripada sistem tanaman bergiliran jagung-kacang tanah dengan aplikasi sumber N yang berbeza. Kajian inkubasi di makmal menggunakan tanah *Ultisol* menunjukkan fluks maksimum N_2O ($2379 \mu g N_2O-N kg^{-1} tanah hari^{-1}$) terjadi apabila ditambah tahi ayam pada 60% ruangrongga isian air (water-filled pore space, WFPS). Penambahan kalium nitrat, sisa kacang tanah dan urea menunjukkan kadar penghasilan N_2O yang rendah ($615 - 669 \mu g N_2O-N kg^{-1} tanah hari^{-1}$). Penambahan ammonium sulfat dan sisa jagung menghasilkan N_2O yang lebih rendah, iaitu masing-masing 229 dan $246 \mu g N_2O-N kg^{-1} tanah hari^{-1}$. Penghasilan jumlah N_2O , dalam 25 hari meningkat dengan penurunan nisbah C/N sumber N organik. Peratus kehilangan baja N melalui penghasilan N_2O adalah lebih

tinggi untuk sumber N tak organik (3.5-8.6%) berbanding sumber N organik (1.6-6.7%), disebabkan berlakunya proses denitrifikasi pada permulaan tempoh inkubasi dengan adanya N mineral tersedia berbanding permineralan N yang berlaku dari baja organik. Walaupun penghasilan N_2O rendah ($26.6\text{--}38.7 \mu\text{g N}_2\text{O-N kg}^{-1} \text{ tanah hari}^{-1}$), fluksnya meningkat selaras dengan peningkatan peratus kelembapan tanah apabila dibandingkan dengan tanah yang lebih kering (20% WFPS) yang bertindak sebagai penjerap N_2O . Penghasilan jumlah N_2O pada 40, 60 dan 80% WFPS masing-masing meningkat sehingga 46, 58 dan 72% pada 20% WFPS. Pengapuran tanah berasid, sama seperti penambahan urea dan tahi ayam, telah meningkatkan pH tanah sehingga 5.5 dan meningkatkan pengumpulan nitrat selepas tempoh lamban dan penghasilan N_2O . Pemeruwapan N_2O tidak dikawal oleh keasidan tanah; jumlah penghasilan N_2O berkorelasi secara positif dengan pH, CEC, C organik dan kandungan N tanah dan berkorelasi negatif dengan karbon organik larut air (WSOC). Dalam sistem tanaman bergiliran jagung-kacang tanah, penambahan tahi ayam menyebabkan fluks N_2O maksimum ($9889 \mu\text{g N}_2\text{O-N kg}^{-1} \text{ tanah m}^{-2} \text{ hari}^{-1}$), dalam masa satu minggu semasa tempoh tanpa tanaman, iaitu selepas tanaman kacang tanah. Kesan sisa tahi ayam dapat dilihat semasa penanaman jagung iaitu dengan kadar pemeruwapan N_2O yang tinggi ($4053 \mu\text{g N}_2\text{O-N kg}^{-1} \text{ tanah hari}^{-1}$). Fluks N_2O yang rendah atau fluks negative semasa tempoh tanpa penanaman terjadi disebabkan substrat yang rendah atau WFPS yang rendah (<40%). Sumber N yang ditambah, kekal di dalam tanah sehingga 2–3 minggu berpadanan dengan pemeruwapan N_2O . Variasi koefisien yang tinggi bagi fluks N_2O untuk kedua-dua tanaman menunjukkan tiada variasi fluks diurnal N_2O yang jelas. Variasi temporal adalah besar, dan menunjukkan pemeruwapan yang tinggi semasa tempoh tanpa tanaman, iaitu selepas penambahan tahi ayam serta semasa penanaman jagung selepas penambahan baja N. Jumlah pemeruwapan N_2O tertinggi dalam tempoh penanaman jagung ($1.82 \text{ kg N}_2\text{O-N ha}^{-1}$), adalah dalam plot penambahan tahi ayam bersama separuh daripada kadar baja N yang disyorkan (75 kg N ha^{-1}), mungkin disebabkan kesan penambahan tahi ayam sebelum penanaman

jagung. Pemeruwapan tahunan yang dicatatkan adalah 3.94, 1.90 dan 1.41 kg N₂O-N ha⁻¹ daripada plot penambahan tahi ayam bersama sisa tanaman dan baja N, plot sisa tanaman bersama baja N, dan plot baja N sahaja. Faktor pemeruwapan N₂O disebabkan penambahan baja (1.06%) yang dikira daripada kajian ini, adalah lebih rendah daripada nilai yang digunakan sekarang mengikut garis panduan 'Intergovernmental Panel on Climate Change' iaitu 1.25%. Keputusan ini menunjukkan bahawa penambahan tahi ayam di kawasan tanaman adalah berpotensi sebagai punca utama pemeruwapan N₂O.

ACKNOWLEDGEMENTS

The author extends his deep sense of gratitude to the Chairman of Supervisory Committee Associate Professor Dr. Rosenani Abu Bakar, Department of Land Mangement, Universiti Putra Malaysia (UPM) and the Co-Chairman of the Supervisory Committee Professor Dr. ir. Oswald Van Cleemput, Faculty of Agricultural and Applied Biological Sciences, Ghent University, Belgium, for their scholastic and active guidance, valuable advice and tutelage during the research work and preparation of his dissertation.

A sincere gratitude also goes to the members of the Supervisory Committee Professor Dr. Shamshuddin Jusop and Dr. Che Fauziah Ishak, Department of Land Management, UPM, for their technical advice and valuable guidance at various stages of research and critical reviewing of his dissertation.

He gratefully acknowledges the Ghent University, Belgium and Universiti Putra Malaysia and the respective Governments for providing the scholarship and research facilities under a Ph.D Twinning Programme. He is indebted to the Bangladesh Institute of Nuclear Agriculture (BINA) and the Government of the Peoples' Republic of Bangladesh for providing deputation and immense help to accomplish his degree.

The author wishes to express his appreciation to Professor Dr. G. Stoops, Co-ordinator of the Twinning Programme and Dr. ir. Pascal Boeckx, Laboratory of Applied Physical Chemistry, Ghent University, Belgium for their assistance and suggestions.

He desires to extend his appreciation to Associate Professor Dr. Siti Zauyah Darus, Dr. Anuar Abdul Rahim, Mr. Peli Mat, Associate Professor Dr. Aminuddin Hussin, Associate Professor Dr. Zaharah Abdul Rahman and Associate Professor Dr. Mohd. Khanif Yusop of UPM for their help and moral supports.

He is profoundly indebted to the technical staff of the Department of Land Management of UPM, especially to Mr. Mutuviren, Mrs. Faridah, Mrs. Rusnah, Mr. Sabri, Mrs. Norhayati, Mr Shukri, Mrs. Fauziah, Mrs. Faridah (MSTB) and other staffs of the Soil Science Laboratories for their co-operation during my research work..

The author is thankful to all of his friends and colleagues of national and international origins, especially to Mr. Amir Hamzah for translating the abstract, and to Dr. M.A. Satter, Mr. M.A. Baset Mia, Mr. Humayun Kabir Mithu, Mr. Shibli Russel for providing an amiable atmosphere and encouragement.

He expresses his high regard to his mother and late father for being the pillar, who inculcated academic perseverance. He is highly grateful to his loving wife, Ferdousi Begum Lucky and sweet daughter, Dimitra Khalil Chaity for their patience, sacrifices and a never-ending inspiration.

I certify that an Examination Committee met on 2nd March 2001 to conduct the final examination of Mohammad Ibrahim Khalil on his Doctor of Philosophy thesis entitled "Nitrous Oxide Emission from an Upland Cropping System in the Humid Tropics" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

WAN SULAIMAN WAN HARUN, Ph.D.

Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

ROSENANI ABU BAKAR, Ph.D.

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

OSWALD VAN CLEEMPUT, Ph.D. Ir.

Professor
Faculty of Agricultural and Applied Biological Sciences
Ghent University, Belgium
(Member)

SHAMSHUDDIN JUSOP, Ph.D.

Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

CHE FAUZIAH ISHAK, Ph.D.

Faculty of Agriculture
Universiti Putra Malaysia
(Member)

TEE BOON GOH, Ph.D.

Professor
Department of Soil Science
University of Manitoba, Canada
(Independent Examiner)



MOHD/CHAZALI MOHAYIDIN, Ph.D.
Professor/Deputy Dean of Graduate School
Universiti Putra Malaysia

Date: 12 MAR 2001

This thesis submitted to the Senate of Universiti Putra Malaysia has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy.

MOHD. GHAZALI MOHAYIDIN, Ph.D.

Professor

Deputy Dean of Graduate School

Universiti Putra Malaysia

Date:

I hereby declare that this thesis is based on my original work except for quotations and citations, which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.



MOHAMMAD IBRAHIM KHALIL

Date: 09/03/2009

TABLE OF CONTENTS

DEDICATION.....	ii
ABSTRACT.....	iii
ABSTRAK.....	vi
ACKNOWLEDGEMENTS.....	ix
APPROVAL SHEETS.....	xi
DECLARATION FORM.....	xiii
LIST OF TABLES.....	xvi
LIST OF FIGURES.....	xvii

CHAPTER

I	INTRODUCTION	1
II	LITERATURE REVIEW	6
	2.1 Nitrous Oxide and Global Climate Change	6
	2.1.1 Global Warming	7
	2.1.2 Ozone Layer Depletion	8
	2.1.3 Acid Rain Formation	10
	2.2 Nitrous Oxide Formation in the Soil System	11
	2.2.1 Sources of N ₂ O and Agriculture	11
	2.2.2 Pathways of N ₂ O Formation from Agricultural Soils	13
	2.3 Processes and Factors Affecting N ₂ O Production/Diffusion in the Soil System	15
	2.3.1 Processes of N ₂ O Production	15
	2.3.2 Factors Affecting N ₂ O Production/Diffusion	18
	2.4 Variability of N ₂ O Emissions from Agroecosystems	38
	2.4.1 Spatial Variations of N ₂ O Emissions	39
	2.4.2 Diurnal Variations of N ₂ O Emissions	40
	2.4.3 Temporal Variations of N ₂ O Emissions	41
	2.4.4 Seasonal Variations of N ₂ O Emissions	44
	2.5 N ₂ O Emissions from Upland Cropping Systems	45
	2.5.1 N ₂ O Emissions from Maize Fields	46
	2.5.2 N ₂ O Emissions from Legume Cover	50
	2.5.3 N ₂ O Emissions from Fallow Lands	52
	2.5.4 N ₂ O Emissions from Other Upland Cropping Systems	53
III	NITROUS OXIDE PRODUCTION AS INFLUENCED BY DIFFERENT NITROGEN SOURCES, MOISTURE REGIMES, LIMING AND SOIL TYPE: A LABORATORY STUDY	57
	3.1 Introduction	57
	3.2 Materials and Methods	59
	3.2.1 Description of Soils	59
	3.2.2 Experimental Treatments	61



3.2.3 Preincubation	63
3.2.4 Experimental Procedures	63
3.2.5 Gas Sampling and Measurement of N ₂ O	64
3.2.6 Soil Analyses	66
3.2.7 Statistical Analyses	68
3.3 Results and Discussion	68
3.3.1 Nitrous Oxide Fluxes	68
3.3.2 Total N ₂ O Production	97
3.4. Conclusions	109
IV NITROUS OXIDE EMISSION FROM A MAIZE- GROUNDNUT CROPPING SYSTEM IN THE HUMID TROPICS	112
4.1 Introduction	112
4.2 Materials and Methods	114
4.2.1 Location and Soil Characteristics	114
4.2.2 Treatments and Experimental Design	115
4.2.3 Inorganic and Organic Fertilizer Applications	115
4.2.4 Sowing and Harvest	116
4.2.5 Standardization of Gas Chamber	116
4.2.6 Installation of Gas Chamber	117
4.2.7 Collection of Gas Samples	119
4.2.8 Analyses of Gas Samples	119
4.2.9 Soil Samplings and Temperature Measurements	121
4.2.10 Soil Analyses	121
4.2.11 Statistical Analyses and Estimation of Variability	122
4.3 Results and Discussion	124
4.3.1 Nitrous Oxide Fluxes	124
4.3.2 Diurnal Variations of N ₂ O Flux	151
4.3.3 Total N ₂ O Emissions	154
4.4 Conclusions	163
V SUMMARY AND CONCLUSIONS	165
REFERENCES	172
APPENDICES	197
VITA	204

LIST OF TABLES

Table		Page
2.1	Global N ₂ O budgets (IPCC, 1997)	12
2.2a	Nitrous oxide emissions from the upland crop fields other than maize and leguminous crops	55
2.2b	Nitrous oxide emissions from the upland crop fields other than maize and leguminous crops	56
3.1	Physical and chemical properties of different soil type	60
3.2	Influence of applied N fertilizers on total N ₂ O production and loss during 25 days of the incubation	98
3.3	Influence of crop residues and chicken manure on total N ₂ O production and loss during 25 days of the incubation	100
3.4	Relationships between total N ₂ O production and some indigenous soil properties as influenced by soil types	108
4.1	Spatial variations of N ₂ O flux as influenced by cropping/fallow periods and treatments	126
4.2	Temporal variations of N ₂ O emission based on the days of higher N ₂ O flux during maize-groundnut cropping system as influenced by inorganic and organic N sources	156

LIST OF FIGURES

Figure		Page
2.1	Nitrogen cycle of agricultural soils and its relationship to N ₂ O production	14
3.1	The N ₂ O flux (a), NH ₄ ⁺ -N (b) and NO ₃ ⁻ -N (c) with time as influenced by different nitrogenous fertilizers during 25 days of the incubation	70
3.2	The water-soluble organic carbon (WSOC) (a), and pH (H ₂ O) (b) with time as influenced by different nitrogenous fertilizers during 25 days of the incubation	72
3.3	The N ₂ O flux (a), NH ₄ ⁺ -N (b) and NO ₃ ⁻ -N (c) with time as influenced by crop residues and chicken manure during 25 days of the incubation	76
3.4	The water-soluble organic carbon (WSOC) (a), and pH _{H2O} (b) with time as influenced by crop residues and chicken manure during 25 days of the incubation	79
3.5	The N ₂ O flux (a), NH ₄ ⁺ -N (b) and NO ₃ ⁻ -N (c) with time as influenced by moisture regimes during 25 days of the incubation	83
3.6	The water-soluble organic carbon (WSOC) (a), and pH _{H2O} (b) with time as influenced by moisture regimes during 25 days of the incubation	86
3.7	The N ₂ O flux (a), NH ₄ ⁺ -N (b) and NO ₃ ⁻ -N (c) with time as influenced by soil types with liming during 40 days of the incubation	89
3.8	The water-soluble organic carbon (WSOC) (a), and pH _{H2O} (b) with time as influenced by soil types with liming during 40 days of the incubation	91
3.9	The N ₂ O flux (a) NH ₄ ⁺ -N (b) and NO ₃ ⁻ -N (c) with time as influenced by soil types without liming during 40 days of the incubation	94
3.10	The water-soluble organic carbon (WSOC) (a), and pH _{H2O} (b) with time as influenced by soil types with liming during 40 days of the incubation	96

3.11	Total N ₂ O emissions as influenced by moisture regimes during 25 days of the incubation	103
3.12	Total N ₂ O emissions as influenced by soil types with liming during 40 days of the incubation	105
3.13	Total N ₂ O emissions as influenced by soil types without liming during 40 days of the incubation	106
4.1	Relationships between N ₂ O concentration and time interval for gas collection as influenced by C and N sources. gas was collected at day 2 (a and c, without and with C source) and at day 6 (b and d, without and with C source, respectively)	118
4.2	A PVC chamber (closed system) with different instruments used for gas collection that placed in between two plants on the ridge of the furrow	120
4.3	Daily minimum and maximum air temperature and rainfall during the groundnut-fallow-maize-fallow period	125
4.4	Nitrous oxide fluxes during groundnut growing period as influenced by inorganic and organic N sources	128
4.5	The NH ₄ ⁺ -N (a), NO ₃ ⁻ -N (b) and NO ₂ ⁻ -N (c) with time during groundnut period as influenced by application of inorganic and organic fertilizers	130
4.6	The soil pH (a), water-soluble organic carbon (WSOC) (b), water-filled pore spaces (WFPS) (c), and soil temperature (d) with time during groundnut period as influenced by application of inorganic and organic fertilizers	131
4.7	Nitrous oxide fluxes during maize growing period as influenced by inorganic and organic N sources	133
4.8	The NH ₄ ⁺ -N (a), NO ₃ ⁻ -N (b) and NO ₂ ⁻ -N (c) with time during the maize period as influenced by application of inorganic and organic fertilizers	135
4.9	The soil pH (a), water-soluble organic carbon (WSOC) (b), water-filled pore spaces (WFPS) (c), and soil temperature (d) with time during the maize period as influenced by application of inorganic and organic fertilizers	136
4.10	Nitrous oxide fluxes during the fallow period after groundnut as influenced by inorganic and organic N sources	139

4.11	The $\text{NH}_4^+\text{-N}$ (a), $\text{NO}_3^-\text{-N}$ (b) and $\text{NO}_2^-\text{-N}$ (c) with time during the fallow period after groundnut as influenced by application of inorganic and organic fertilizers	141
4.12	The soil pH (a), water-soluble organic carbon (WSOC) (b), water-filled pore spaces (WFPS) (c), and soil temperature (d) with time during the fallow period after groundnut as influenced by application of inorganic and organic fertilizers	142
4.13	Nitrous oxide fluxes during the fallow period after maize as influenced by inorganic and organic N sources	145
4.14	The $\text{NH}_4^+\text{-N}$ (a), $\text{NO}_3^-\text{-N}$ (b) and $\text{NO}_2^-\text{-N}$ (c) with time during the fallow period after maize as influenced by application of inorganic and organic fertilizers	147
4.15	The soil pH (a), water-soluble organic carbon (WSOC) (b), water-filled pore spaces (WFPS) (c), and soil temperature (d) with time during the fallow period after maize as influenced by application of inorganic and organic fertilizers	148
4.16	Nitrous oxide fluxes during the maize-groundnut rotation system as influenced by inorganic and organic N sources	150
4.17	Diurnal variations of N_2O fluxes and changes in soil and air temperature measured during the gas collection period in each treatment plots under groundnut cover	152
4.18	Diurnal variations of N_2O fluxes and changes in soil and air temperature measured during the gas collection period in each treatment plots under maize cover	152
4.19	Relative deviation from the annual mean (RDAM) of total N_2O emission (solid line) as affected by inorganic and organic N sources during maize-groundnut crop rotation and monthly rainfall	157
4.20	The total N_2O emission during groundnut, maize and fallow periods as influenced by inorganic and organic N sources	160
4.21	Annual N_2O emission as influenced by inorganic and organic N sources during maize-groundnut crop rotation	162

CHAPTER I

INTRODUCTION

Nitrogen, an essential element for plant growth, plays a vital role in the soil-plant-atmosphere continuum. It is estimated that, by the year 2020 at a global level, 70% of the plant nutrients will have to come from fertilizers with a view to sustain the future world population (Ayoub, 1999). The annual global use of fertilizers will need to be doubled by the year 2030 from about 130 million tonnes in the 1990s (Brown et al., 1997), if the current per capita cereal production is to be maintained (Gilland, 1993). Besides, the anthropogenic N inputs into agricultural systems like N from animal wastes, increased biological N fixation, cultivation of mineral and organic soils and addition of crop residue to the field are also a growing concern. The use of inorganic nitrogenous fertilizers has been increasing in the tropics during the last few decades to enhance soil productivity and crop yield potential. Consequently, the indiscriminate use of both inorganic and organic N fertilizers may cause significantly higher gaseous N losses, particularly nitrous oxide (N_2O) that causes global warming and ozone layer depletion (Bouwman, 1990; Cicerone, 1987; Crutzen, 1981). The main sources of N_2O are cultivated soils, biomass burning, fossil fuels and nitric and adipic acid production. On a molar basis, N_2O is about 250-320 times more effective as an absorber of infrared radiation than CO_2 (IPCC, 1995; Robertson, 1993) and about 25 times more than CH_4 (Murdiyarso, 1998). The atmospheric concentrations of N_2O have increased by 15%

during the last 250 years (Mosier, 1998). The present increasing concentration of N_2O in the atmosphere seems to create a genuine catastrophe on the global climate.

The N_2O emission is a significant biogenic phenomenon in N transformation mechanisms and occurs during both the nitrification and denitrification process. It may be formed by various denitrifiers, nitrifiers and even certain assimilatory nitrate-reducing yeasts and fungi. Nitrification may be a significant source of N_2O through autotrophic microbes in most soils and heterotrophs in aerobic to near-aerobic soils, particularly in soils that are too acidic to support the chemoautotrophic nitrifiers (Anderson et al., 1993; Bremner, 1997). Its production is enhanced in soils having a high mineralization capacity to form NH_4^+ or treated with nitrifiable forms of nitrogen. The N_2O is an obligatory intermediate during denitrification and aerobic bacteria are basically responsible for the process. The dominant denitrifiers are organotrophs because of their versatility and ability to compete for C substrate (Tiedje, 1988). If soils containing nitrate become anaerobic, the availability of organic carbon to enhance the activity of denitrifiers is the limiting factor for the reduction of nitrate. During both processes, a large accumulation of NO_2^- -N can be a key compound in N loss processes, forming NO, NO_2 and N_2O (Firestone and Davidson, 1989) because of its low stability in acid conditions (Van Cleemput and Baert, 1984).

The emission of N_2O to the atmosphere from the soil system consists of a series of complex reactions. It is also related to the sequence of enzymatic processes in which the living microbial biomass provides the enzymes and the dead microbial biomass the substrate (Mengel, 1996). The N_2O release depends on the N supplying capacity of

soils, which depends mostly on the indigenous soil organic matter, addition of organic residue and the various soil environmental factors viz. moisture, temperature, aeration and pH (Németh and Szebeni, 1987; Szebeni and Németh, 1987). Under aerobic conditions, nitrification is the dominant process for N_2O formation, though a small uptake has been observed in isolated instances in dry soils (Duxbury and Mosier, 1993). It is greater under anaerobic conditions (Firestone, 1982) during denitrification. However, its consumption has also been reported in wet grass pastures (Ryden, 1981). Its production and diffusion are considerable upon irrigation/rainfall events under upland conditions by changing the soil physico-chemical properties or by affecting soil gas diffusivity and microbial activity and subsequent nitrogen gas production and efflux (Delgado and Mosier, 1996; Valente and Thornton, 1993). However, Rosswall et al. (1989) emphasized on the medium to high moisture content, limiting oxygen diffusion, and high mineral-N and high organic-C availability for the production of N_2O from soils.

The application of chemical N fertilizers is a major contribution to N_2O emission from agricultural soils. Addition of organic residue/amendment, preferably N-rich residue, causes considerable release of N_2O . It is estimated that more than 75% of the added N fertilizer is lost from the residue-soil system on a year to year basis if the soil N content remains unchanged (Beauchamp, 1997). In general, N_2O emissions from agricultural land vary from 0.03 to 2.7% of the applied total N fertilizer (Eichner, 1990). However, soil management and cropping systems, and variable rainfall have greater effects on N_2O emission than the type of fertilizer and its fluxes are variable in time and space (Mosier, 1989). Biological nitrogen fixation (BNF) also acts as a source

of N_2O as the atmospheric nitrogen fixed by legumes can be nitrified and denitrified in the same way as fertilizer nitrogen (Freney, 1997; Galbally et al., 1992). The contribution from the BNF ranges from 0.5 to 5 kg $\text{N}_2\text{O-N ha}^{-1} \text{ yr}^{-1}$ that vary with soils and climatic conditions (Carran et al., 1995; Mosier et al., 1996). However, the fixation and conversion coefficients are still uncertain.

Ultisols, Oxisols and Inceptisols are the dominant soils in Malaysia, where Oxisols and Ultisols occupy about 72% of the total area. They are also the major soils of the tropics and occupy 38.1% of the total land area, where the Ultisols covers 10.6% (Van Wambeke, 1991). Malaysian soils in the upland are mostly weathered and acid with low organic matter content and low CEC. Hence, more and more inorganic and organic N fertilizers are applied to sustain yields through improvement of soil productivity. As a typical humid country in the tropics, this area mostly experiences a good amount of rainfall (2000-2500 mm per annum), which is well distributed, and has a temperature range of 24-34°C throughout the year. These may influence gaseous and leaching losses of N with poor N use efficiency by the crops. Controlled release fertilizer or nitrification inhibitor has the potential to improve N use efficiency by matching nutrient release with crop demand and reducing nitrate release and gaseous losses (Delgado and Mosier, 1996). However, its application is still considered to be uneconomical due to the higher production cost of the fertilizers. Appropriate soil management approaches may be considered better options till now to reduce the emission of N_2O gas - a global concern for the 21st century.

Objectives of the Study

Identification of the processes involved in N_2O production from agricultural systems may take into account also different soils, crops and climates. However, research work has mostly been confined on its emission in the temperate regions. There is only limited information concerning utilization of N from crop residue and animal manure applied to agricultural soils (Mosier et al., 1998a). In the humid tropics, considerable works on rice-ecosystem has been done and very few on the upland agroecosystems, particularly in acid soils. Maize is one of the major crops in the tropics, next to rice and wheat. Groundnut, a leguminous oil crop, occupies a large area next to oil palm, soybean and mustard and has also been cultivated either as monocrop or in rotation. However, information on N_2O emission from a maize-groundnut crop rotation is greatly lacking, particularly under sustainable soil management systems. Therefore, this study was carried out to estimate the emission of N_2O from an upland cropping system applied with both inorganic and organic N (as crop residue/amendment) fertilizers. The following specific objectives are defined:

1. To study the diurnal and temporal variations of N_2O emission, and the annual N_2O release from a maize-groundnut crop rotation under different soil management practices over a one-cycle period.
2. To measure N_2O fluxes under different inorganic and organic nitrogenous fertilizers, and moisture regimes through the laboratory incubation technique using the soil of the experimental field.
3. To evaluate N_2O production under laboratory conditions using different soil types with or without liming and to determine soil factors controlling its production.